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DESCRIPTION**METHOD FOR DIAGNOSING CHRONIC MYELOID LEUKEMIA
FIELD OF THE INVENTION**

The invention relates to methods of diagnosing chronic myeloid leukemia.

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PRIORITY INFORMATION

This application claims priority to United States Provisional Application Serial No.60/414,867, filed September 30, 2002.

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BACKGROUND OF THE INVENTION

Chronic myeloid leukemia (CML) is a clonal myeloproliferative disease characterized by Philadelphia (Ph) chromosome translocation (1). The resulting BCR-ABL fusion gene encodes a cytoplasmic protein that is constitutively activated for its tyrosine kinase activity. CML progresses through distinct clinical stages; the earliest stage, termed the chronic phase, is characterized by expansion of terminally differentiated neutrophils. The acute phase termed accelerated phase and blast crisis characterized by maturation arrest with excessive numbers of undifferentiated myeloid or lymphoid progenitor cells (2). Current therapies include allogenic stem-cell transplantation (SCT) and chemotherapies including interferon- α (IFN- α) (3). IFN- α prolongs overall survival but has considerable adverse effects. SCT is the only curative treatment, but is associated with substantial morbidity and is limited to patients with suitable donors.

The development of the ABL-selective tyrosine kinase inhibitor STI571 (imatinib; Glivec; Novartis, Basel, Switzerland) is a significant advance in the management of CML (4,5). STI571 frequently induces remarkable hematologic and cytogenetic responses in these clinical settings. However, recent clinical studies with STI571 in CML demonstrated that many patients at the advanced stage respond well but then relapse (6),(7). Resistance to STI571 because of enhanced expression or mutation of BCR-ABL gene has been found in CML patients (8), (9). Indeed, STI571-induced hematologic responses occur less frequently and are less durable in CML patients at the blast crisis phase.

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cDNA microarray technologies have enabled to obtain comprehensive profiles of gene expression in normal and malignant cells, and compare the gene expression in malignant and corresponding normal cells (Okabe et al., Cancer Res 61:2129-37 (2001); Kitahara et al., Cancer Res 61: 3544-9 (2001); Lin et al., Oncogene 21:4120-8 (2002); Hasegawa et al., Cancer Res 5 62:7012-7 (2002)). This approach enables to disclose the complex nature of cancer cells, and helps to understand the mechanism of carcinogenesis. Identification of genes that are deregulated in tumors can lead to more precise and accurate diagnosis of individual cancers, and to develop novel therapeutic targets (Bienz and Clevers, Cell 103:311-20 (2000)). To disclose mechanisms underlying tumors from a genome-wide point of view, and discover target 10 molecules for diagnosis and development of novel therapeutic drugs, the present inventors have been analyzing the expression profiles of tumor cells using a cDNA microarray of 23040 genes (Okabe et al., Cancer Res 61:2129-37 (2001); Kitahara et al., Cancer Res 61:3544-9 (2001); Lin et al., Oncogene 21:4120-8 (2002); Hasegawa et al., Cancer Res 62:7012-7 (2002)).

Studies designed to reveal mechanisms of carcinogenesis have already facilitated 15 identification of molecular targets for anti-tumor agents. For example, inhibitors of farnesyltransferase (FTIs) which were originally developed to inhibit the growth-signaling pathway related to Ras, whose activation depends on posttranslational farnesylation, has been effective in treating Ras-dependent tumors in animal models (He et al., Cell 99:335-45 (1999)). Clinical trials on human using a combination of anti-cancer drugs and anti-HER2 monoclonal 20 antibody, trastuzumab, have been conducted to antagonize the proto-oncogene receptor HER2/neu; and have been achieving improved clinical response and overall survival of breast-cancer patients (Lin et al., Cancer Res 61:6345-9 (2001)). A tyrosine kinase inhibitor, STI-571, which selectively inactivates bcr-abl fusion proteins, has been developed to treat chronic myelogenous leukemias wherein constitutive activation of bcr-abl tyrosine kinase plays a crucial 25 role in the transformation of leukocytes. Agents of these kinds are designed to suppress oncogenic activity of specific gene products (Fujita et al., Cancer Res 61:7722-6 (2001)). Therefore, gene products commonly up-regulated in cancerous cells may serve as potential targets for developing novel anti-cancer agents.

It has been demonstrated that CD8+ cytotoxic T lymphocytes (CTLs) recognize epitope 30 peptides derived from tumor-associated antigens (TAAs) presented on MHC Class I molecule, and lyse tumor cells. Since the discovery of MAGE family as the first example of TAAs, many other TAAs have been discovered using immunological approaches (Boon, Int J Cancer 54: 177-

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80 (1993); Boon and van der Bruggen, *J Exp Med* 183: 725-9 (1996); van der Bruggen et al., *Science* 254: 1643-7 (1991); Brichard et al., *J Exp Med* 178: 489-95 (1993); Kawakami et al., *J Exp Med* 180: 347-52 (1994)). Some of the discovered TAAs are now in the stage of clinical development as targets of immunotherapy. TAAs discovered so far include MAGE (van der
5 Bruggen et al., *Science* 254: 1643-7 (1991)), gp100 (Kawakami et al., *J Exp Med* 180: 347-52 (1994)), SART (Shichijo et al., *J Exp Med* 187: 277-88 (1998)), and NY-ESO-1 (Chen et al., *Proc Natl Acad Sci USA* 94: 1914-8 (1997)). On the other hand, gene products which had been demonstrated to be specifically overexpressed in tumor cells, have been shown to be recognized as targets inducing cellular immune responses. Such gene products include p53 (Umano et al.,
10 *Brit J Cancer* 84: 1052-7 (2001)), HER2/neu (Tanaka et al., *Brit J Cancer* 84: 94-9 (2001)), CEA (Nukaya et al., *Int J Cancer* 80: 92-7 (1999)), and so on.

In spite of significant progress in basic and clinical research concerning TAAs (Rosenbeg et al., *Nature Med* 4: 321-7 (1998); Mukherji et al., *Proc Natl Acad Sci USA* 92: 8078-82 (1995); Hu et al., *Cancer Res* 56: 2479-83 (1996)), only limited number of candidate TAAs for the
15 treatment of adenocarcinomas, including colorectal cancer, are available. TAAs abundantly expressed in cancer cells, and at the same time which expression is restricted to cancer cells would be promising candidates as immunotherapeutic targets. Further, identification of new TAAs inducing potent and specific antitumor immune responses is expected to encourage clinical use of peptide vaccination strategy in various types of cancer (Boon and van der Bruggen,
20 *J Exp Med* 183: 725-9 (1996); van der Bruggen et al., *Science* 254: 1643-7 (1991); Brichard et al., *J Exp Med* 178: 489-95 (1993); Kawakami et al., *J Exp Med* 180: 347-52 (1994); Shichijo et al., *J Exp Med* 187: 277-88 (1998); Chen et al., *Proc Natl Acad Sci USA* 94: 1914-8 (1997); Harris, *J Natl Cancer Inst* 88: 1442-5 (1996); Butterfield et al., *Cancer Res* 59: 3134-42 (1999); Vissers et al., *Cancer Res* 59: 5554-9 (1999); van der Burg et al., *J Immunol* 156: 3308-14
25 (1996); Tanaka et al., *Cancer Res* 57: 4465-8 (1997); Fujie et al., *Int J Cancer* 80: 169-72 (1999); Kikuchi et al., *Int J Cancer* 81: 459-66 (1999); Oiso et al., *Int J Cancer* 81: 387-94 (1999)).

It has been repeatedly reported that peptide-stimulated peripheral blood mononuclear cells (PBMCs) from certain healthy donors produce significant levels of IFN- γ in response to the peptide, but rarely exert cytotoxicity against tumor cells in an HLA-A24 or -A0201 restricted
30 manner in ⁵¹Cr-release assays (Kawano et al., *Cancer Res* 60: 3550-8 (2000); Nishizaka et al., *Cancer Res* 60: 4830-7 (2000); Tamura et al., *Jpn J Cancer Res* 92: 762-7 (2001)). However, both of HLA-A24 and HLA-A0201 are one of the popular HLA alleles in Japanese, as well as

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Caucasian (Date et al., Tissue Antigens 47: 93-101 (1996); Kondo et al., J Immunol 155: 4307-12 (1995); Kubo et al., J Immunol 152: 3913-24 (1994); Imanishi et al., Proceeding of the eleventh International Hictocompatibility Workshop and Conference Oxford University Press, Oxford, 1065 (1992); Williams et al., Tissue Antigen 49: 129 (1997)). Thus, antigenic peptides
5 of carcinomas presented by these HLAs may be especially useful for the treatment of carcinomas among Japanese and Caucasian. Further, it is known that the induction of low-affinity CTL in vitro usually results from the use of peptide at a high concentration, generating a high level of specific peptide/MHC complexes on antigen presenting cells (APCs), which will effectively activate these CTL (Alexander-Miller et al., Proc Natl Acad Sci USA 93: 4102-7 (1996)).

SUMMARY OF THE INVENTION

The invention is based upon the discovery of a pattern of gene expression correlated with CML. The genes that are differentially expressed in CML are collectively referred to herein as "CML nucleic acids" or "CML polynucleotides" and the corresponding encoded polypeptides are referred to as "CML polypeptides" or "CML proteins."

15 Accordingly, the invention features a method of diagnosing or determining a predisposition to CML in a subject by determining an expression level of a CML-associated gene in a patient derived biological sample, such as peripheral blood sample or myeloid cells sample. By CML associated gene is meant a gene that is characterized by an expression level which differs in a cell containing a Philadelphia (Ph) chromosome translocation or in a cell obtained
20 from an individual with a family history of CML or an individual exhibiting clinical symptoms of CML, compared to a normal peripheral blood cell. A CML-associated gene is one or more of CML 1-296. An alteration, *e.g.*, increase or decrease of the level of expression of the gene compared to a normal control level of the gene indicates that the subject suffers from or is at risk of developing CML.

25 By normal control level is meant a level of gene expression detected in a normal, healthy individual or in a population of individuals known not to be suffering from CML. A control level is a single expression pattern derived from a single reference population or from a plurality of expression patterns. For example, the control level can be a database of expression patterns from previously tested cells. A normal individual is one with no clinical symptoms of CML and
30 no family history of CML.

An increase in the level of CML 1-190 detected in a test sample compared to a normal

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control level indicates the subject (from which the sample was obtained) suffers from or is at risk of developing CML. In contrast, a decrease in the level of CML 191-296 detected in a test sample compared to a normal control level indicates said subject suffers from or is at risk of developing CML.

5 Alternatively, expression of a panel of CML-associated genes in the sample is compared to a CML control level of the same panel of genes. By CML control level is meant the expression profile of the CML-associated genes found in a population suffering from CML.

Gene expression is increased or decreased 10%, 25%, 50% compared to the control level. Alternately, gene expression is increased or decreased 1, 2, 5 or more fold compared to the
10 control level. Expression is determined by detecting hybridization, *e.g.*, on an array, of a CML-associated gene probe to a gene transcript of the patient-derived cell sample.

The patient-derived cell sample is any cell from a test subject, *e.g.*, a patient known to or suspected of having CML. For example, the sample contains a mixture of mononuclear cells from peripheral blood.

15 The invention also provides a CML reference expression profile of a gene expression level of two or more of CML 1-296. Alternatively, the invention provides a CML reference expression profile of the levels of expression of two or more of CML 1-190 or CML 191-296.

The invention further provides methods of identifying an agent that inhibits or enhances the expression or activity of a CML-associated gene, *e.g.* CML 1-296 by contacting a test cell
20 expressing a CML associated gene with a test agent and determining the expression level of the CML associated gene. The test cell is a mononuclear cell such as a mononuclear cell from peripheral blood of CML patient. A decrease of the level compared to a normal control level of the gene indicates that the test agent is an inhibitor of the CML-associated gene and reduces a symptom of CML, *e.g.*, CML 1-190. Alternatively, an increase of the level or activity compared
25 to a normal control level or activity of the gene indicates that said test agent is an enhancer of expression or function of the CML-associated gene and reduces a symptom of CML, *e.g.*, CML 191-296.

The invention also provides a kit with a detection reagent which binds to two or more CML nucleic acid sequences or which binds to a gene product encoded by the nucleic acid
30 sequences. Also provided is an array of nucleic acids that binds to two or more CML nucleic acids.

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Therapeutic methods include a method of treating or preventing CML in a subject by administering to the subject an antisense composition. The antisense composition reduces the expression of a specific target gene, e.g., the antisense composition contains a nucleotide, which is complementary to a sequence selected from the group consisting of CML 1-190. Another method includes the steps of administering to a subject an short interfering RNA (siRNA) composition. The siRNA composition reduces the expression of a nucleic acid selected from the group consisting of CML 1-190. In yet another method, treatment or prevention of CML in a subject is carried out by administering to a subject a ribozyme composition. The nucleic acid-specific ribozyme composition reduces the expression of a nucleic acid selected from the group consisting of CML 1-190. Other therapeutic methods include those in which a subject is administered a compound that increases the expression of CML 191-296 or activity of a polypeptide encoded by CML 191-296. Furthermore, CML can be treated by administering a protein encoded by CML 191-296. The protein may be directly administered to the patient or, alternatively, may be expressed in vivo subsequent to being introduced into the patient, for example, by administering an expression vector or host cell carrying the down-regulated marker gene of interest. Suitable mechanisms for in vivo expression of a gene of interest are known in the art.

The invention also includes vaccines and vaccination methods. For example, a method of treating or preventing CML in a subject is carried out by administering to the subject a vaccine containing a polypeptide encoded by a nucleic acid selected from the group consisting of CML 1-190 or an immunologically active fragment such a polypeptide. An immunologically active fragment is a polypeptide that is shorter in length than the full-length naturally-occurring protein and which induces an immune response. For example, an immunologically active fragment at least 8 residues in length and stimulates an immune cell such as a T cell or a B cell. Immune cell stimulation is measured by detecting cell proliferation, elaboration of cytokines (e.g., IL-2), or production of an antibody.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present

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specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

One advantage of the methods described herein is that the disease is identified prior to detection of overt clinical symptoms such as expansion of terminally-differentiated neutrophils.

5 Other features and advantages of the invention will be apparent from the following detailed description, and from the claims.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a photograph of a DNA agarose gel showing expression of representative 11 genes and β -actin examined by semi-quantitative RT-PCR using cDNA prepared from 10 amplified RNA. The first lane shows the expression level of each gene in a normal individual. The remaining eight lanes each show the expression level of the genes in a different CML patient. Gene symbols are noted for genes whose function was known or inferred and Accession No. Noted for ESTs.

DETAILED DESCRIPTION

15 The present invention is based in part on the discovery of changes in expression patterns of multiple nucleic acid sequences in mononuclear cells from peripheral blood of patients with Chronic myeloid leukemia (CML). The differences in gene expression were identified by using a comprehensive cDNA microarray system.

Using a cDNA microarray containing 23,040 genes, comprehensive gene-expression 20 profiles were obtained of 27 CMLs. Two hundred ninety-six genes were found to be differentially expressed in mononuclear cells from peripheral blood. Results show that certain genes are expressed at low or high levels in CMLs. Selection was made of candidate molecular markers with the potential of detecting CML-related proteins in blood, serum, or sputum of patients, and discovered some potential targets for development of signal- 25 suppressing strategies in CML.

The differentially expressed genes identified herein are used for diagnostic purposes as markers of CML as gene targets, the expression of which is altered to treat or alleviate a symptom of CML.

The genes whose expression levels are modulated (*i.e.*, increased or decreased) in CML

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patients are summarized in Tables 3-4 and are collectively referred to herein as "CML-associated genes", "CML nucleic acids" or "CML polynucleotides" and the corresponding encoded polypeptides are referred to as "CML polypeptides", or "CML proteins." Unless indicated otherwise, "CML" is meant to refer to any of the sequences disclosed herein. (*e.g.*, CML 1-296).

5 The genes that have been previously described are presented along with a database accession number.

By measuring expression of the various genes in a sample of cells, CML is diagnosed. Similarly, by measuring the expression of these genes in response to various agents can identify agents for treating CML.

10 The invention involves determining (*e.g.*, measuring) the expression of at least one, and up to all the CML sequences listed in Tables 3-4. Using sequence information provided by the GeneBank™ database entries for the known sequences the CML associated genes are detected and measured using techniques well known to one of ordinary skill in the art. For example, sequences within the sequence database entries corresponding to CML sequences, are used to
15 construct probes for detecting CML RNA sequences in, *e.g.*, northern blot hybridization analyses. Probes include at least 10, 20, 50, 100, 200 nucleotides of a reference sequence. As another example, the sequences can be used to construct primers for specifically amplifying the CML sequences in, *e.g.*, amplification-based detection methods such as reverse-transcription based polymerase chain reaction.

20 Expression level of one or more of the CML sequences in the test cell population, *e.g.*, a patient derived cell sample is then compared to expression levels of the same sequences in a reference population. The reference cell population includes one or more cells for which the compared parameter is known, *i.e.*, CML cells or non-CML cells.

Whether or not a pattern of gene expression in the test cell population compared to the
25 reference cell population indicates CML or a predisposition thereto depends upon on the composition of the reference cell population. For example, if the reference cell population is composed of non-CML cells, a similar gene expression pattern in the test cell population and reference cell population indicates the test cell population is non-CML. Conversely, if the reference cell population is made up of CML cells, a similar gene expression profile between the
30 test cell population and the reference cell population indicates that the test cell population includes CML cells.

A level of expression of a CML marker gene in a test cell population is considered altered in levels of expression if its expression level varies from the reference cell population by more than 1.0, 1.5, 2.0, 5.0, 10.0 or more fold from the expression level of the corresponding CML sequence in the reference cell population.

5 Differential gene expression between a test cell population and a reference cell population is normalized to a control nucleic acid, *e.g.* a housekeeping gene. For example, a control nucleic acid is one which is known not to differ depending on the CML or non-CML state of the cell. Expression levels of the control nucleic acid in the test and reference nucleic acid can be used to normalize signal levels in the compared populations. Control genes include
10 β -actin, glyceraldehyde 3-phosphate dehydrogenase or ribosomal protein P1.

The test cell population is compared to multiple reference cell populations. Each of the multiple reference populations may differ in the known parameter. Thus, a test cell population may be compared to a second reference cell population known to contain, *e.g.*, CML cells, as well as a second reference population known to contain, *e.g.*, non-CML cells (normal cells). The
15 test cell is included in a tissue type or cell sample from a subject known to contain, or to be suspected of containing, CML cells.

The test cell is obtained from a bodily tissue or a bodily fluid, *e.g.*, biological fluid (such as blood or sputum). For example, the test cell is purified from a tissue. Preferably, the test cell population comprises a mononuclear cell.

20 Cells in the reference cell population are derived from a tissue type similar to test cell. Optionally, the reference cell population is a cell line, *e.g.* a CML cell line (positive control) or a normal non-CML cell line (negative control). Alternatively, the control cell population is derived from a database of molecular information derived from cells for which the assayed parameter or condition is known.

25 The subject is preferably a mammal. The mammal can be, *e.g.*, a human, non-human primate, mouse, rat, dog, cat, horse, or cow.

Expression of the genes disclosed herein is determined at the RNA level using any method known in the art. For example, Northern hybridization analysis using probes which specifically recognize one or more of these sequences can be used to determine gene expression.
30 Alternatively, expression is measured using reverse-transcription-based PCR assays, *e.g.*, using primers specific for the differentially expressed sequences. Expression is also determined at the protein level, *i.e.*, by measuring the levels of polypeptides encoded by the gene products

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described herein, or biological activity thereof. Such methods are well known in the art and include, e.g., immunoassays based on antibodies to proteins encoded by the genes. The biological activity of the proteins encoded by the genes are also well known.

Diagnosing CML

5 CML is diagnosed by measuring the level of expression of one or more CML nucleic acid sequences from a test population of cells, (*i.e.*, a patient derived biological sample). Preferably, the test cell population contains a mononuclear cell, e.g., a cell obtained from peripheral blood. Other biological samples can be used for measuring the protein level. For example, the protein level in the blood, or serum, derived from subject to be diagnosed can be measured by
10 immunoassay or biological assay.

Expression of one or more of CML-associated genes, e.g., CML 1-296 is determined in the test cell or biological sample and compared to the expression of the normal control level. A normal control level is an expression profile of a CML-associated gene typically found in a population known not to be suffering from CML. An increase or a decrease of the level of
15 expression in the patient derived cell sample of the CML-associated genes indicates that the subject is suffering from or is at risk of developing CML. For example, an increase in expression of CML 1-190 in the test population compared to the normal control level indicates that the subject is suffering from or is at risk of developing CML. Conversely, a decrease in expression of CML 191-296 in the test population compared to the normal control level indicates
20 that the subject is suffering from or is at risk of developing CML.

When one or more of the CML-associated genes are altered in the test population compared to the normal control level indicates that the subject suffers from or is at risk of developing CML. For example, at least 1%, 5%, 25%, 50%, 60%, 80%, 90% or more of the panel of CML-associated genes (CML 1-190, CML 191-296, or CML 1-296) are altered.

25

Identifying Agents that inhibit or enhance CML-associated gene expression

An agent that inhibits the expression or activity of a CML-associated gene is identified by contacting a test cell population expressing a CML associated up-regulated gene with a test agent and determining the expression level of the CML associated gene. A decrease in
30 expression in the presence of the agent compared to the normal control level (or compared to the level in the absence of the test agent) indicates the agent is an inhibitor of a CML associated up-regulated gene and useful to inhibit CML.

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Alternatively, an agent that enhances the expression or activity of a CML down-regulated associated gene is identified by contacting a test cell population expressing an CML associated gene with a test agent and determining the expression level or activity of the CML associated down-regulated gene. An increase of expression or activity compared to a normal control level or activity of the CML-associated gene indicates that the test agent augments expression or activity of the down-regulated CML associated gene.

The test cell population is any cell expressing the CML-associated genes. For example, the test cell population contains a mononuclear cell, such a cell is isolated from peripheral blood. Alternatively, the test cell is a cell, which has been transfected with a CML associated gene or which has been transfected with a regulatory sequence (*e.g.* promoter sequence) from a CML-associated gene operably linked to a reporter gene.

Assessing efficacy of treatment of CML in a subject

The differentially expressed CML sequences identified herein also allow for the course of treatment of CML to be monitored. In this method, a test cell population is provided from a subject undergoing treatment for CML. If desired, test cell populations are obtained from the subject at various time points before, during, or after treatment. Expression of one or more of the CML sequences, in the cell population is then determined and compared to a reference cell population which includes cells whose CML state is known. The reference cells have not been exposed to the treatment.

If the reference cell population contains no CML cells, a similarity in expression between CML sequences in the test cell population and the reference cell population indicates that the treatment is efficacious. However, a difference in expression between CML sequences in the test population and a normal control reference cell population indicates a less favorable clinical outcome or prognosis.

By "efficacious" is meant that the treatment leads to a reduction in expression of a pathologically upregulated gene, increase in expression of a pathologically down-regulated gene or a decrease leukemic stem cells and their dividing progeny (*ie.*, granulocytic, erythroid, and megakaryocytic precursors) in a subject. When treatment is applied prophylactically, "efficacious" means that the treatment retards or prevents a symptom of clinical CML. For example, the treatment inhibits a symptom of chronic, acute, or accelerated phase. Assessment of CML is made using standard clinical protocols.

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Efficaciousness is determined in association with any known method for diagnosing or treating CML. CML is diagnosed for example, by identifying symptomatic anomalies, *e.g.*, anemia, hypermetabolism, easy fatigability, weakness, weight loss, and anorexia. Other characteristics of CML include splenomegaly, thrombocytosis and an almost total lack of alkaline phosphatase in granulocytes. Patients also exhibit marked elevation of the leukocyte count with the circulating cells being predominantly neutrophils and metamyelocytes, but basophils and eosinophils may also be prominent. Furthermore, the Ph¹ (Philadelphia) chromosome is present in the dividing progeny of multipotent myeloid stem cells (*ie.*, granulocytic, erythroid, and megakaryocytic precursors) and lymphoid cells (*ie.*, B cells) of approximately 90% of patients with CML.

Selecting a therapeutic agent for treating CML that is appropriate for a particular individual

Differences in the genetic makeup of individuals can result in differences in their relative abilities to metabolize various drugs. An agent that is metabolized in a subject to act as an anti-CML agent can manifest itself by inducing a change in gene expression pattern in the subject's cells from that characteristic of a CML state to a gene expression pattern characteristic of a non-CML state. Accordingly, the differentially expressed CML sequences disclosed herein allow for a putative therapeutic or prophylactic inhibitor of CML to be tested in a test cell population from a selected subject in order to determine if the agent is a suitable anti-CML agent in the subject.

To identify an inhibitor or enhancer of CML, that is appropriate for a specific subject, a test cell population from the subject is exposed to a therapeutic agent, and the expression of one or more of CML 1-296 sequences is determined.

The test cell population contains an CML cell expressing an CML associated gene. Preferably, the test cell is a mononuclear cell. For example a test cell population is incubated in the presence of a candidate agent and the pattern of gene expression of the test sample is measured and compared to one or more reference profiles, *e.g.*, a CML reference expression profile or a non-CML reference expression profile.

A decrease in expression of one or more of the sequences CML 1-190 or an increase in expression of one or more of the sequences CML 191-296 in a test cell population relative to a reference cell population containing CML is indicative that the agent is therapeutic.

The test agent can be any compound or composition. For example, the test agents are agents that regulate growth and differentiation of hematopoietic precursors.

Screening assays for identifying therapeutic agents

5 The differentially expressed sequences disclosed herein can also be used to identify candidate therapeutic agents for treating a CML. The method is based on screening a candidate therapeutic agent to determine if it converts an expression profile of CML 1-296 sequences characteristic of a CML state to a pattern indicative of a non-CML state.

10 In the method, a cell is exposed to a test agent or a combination of test agents (sequentially or consequentially) and the expression of one or more CML 1-296 sequences in the cell is measured. The expression profile of the CML sequences in the test population is compared to expression level of the CML sequences in a reference cell population that is not exposed to the test agent.

15 An agent effective in stimulating expression of underexpressed genes, or in suppressing expression of overexpressed genes is deemed to lead to a clinical benefit such compounds are further tested for the ability to prevent an increased myeloid stem cell mass or to prevent maturation of leukemic stem cells (*ie.*, pluripotent hematopoietic stem cells), in animals or test subjects.

20 In a further embodiment, the present invention provides methods for screening candidate agents which are potential targets in the treatment of CML. As discussed in detail above, by controlling the expression levels or activities of marker genes, one can control the onset and progression of CML. Thus, candidate agents, which are potential targets in the treatment of CML, can be identified through screenings that use the expression levels and activities of marker genes as indices. In the context of the present invention, such screening may comprise, for
25 example, the following steps:

- a) contacting a test compound with a polypeptide encoded by CML 1-296;
- b) detecting the binding activity between the polypeptide and the test compound; and
- c) selecting a compound that binds to the polypeptide

30 Alternatively, the screening method of the present invention may comprise the following steps:

- a) contacting a candidate compound with a cell expressing one or more marker genes, wherein the one or more marker genes is selected from the group consisting of CML 1-

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296; and

- b) selecting a compound that reduces the expression level of one or more marker genes selected from the group consisting of CML 1-190, or elevates the expression level of one or more marker genes selected from the group consisting of CML 191-296.

5 Cells expressing a marker gene include, for example, cell lines established from CML; such cells can be used for the above screening of the present invention.

Alternatively, the screening method of the present invention may comprise the following steps:

- a) contacting a test compound with a polypeptide encoded by selected from the group
10 consisting of CML 1-296;
- b) detecting the biological activity of the polypeptide of step (a); and
- c) selecting a compound that suppresses the biological activity of the polypeptide encoded
by CML 1-190 in comparison with the biological activity detected in the absence of the
test compound, or enhances the biological activity of the polypeptide encoded by CML
15 191-296 in comparison with the biological activity detected in the absence of the test
compound.

A protein required for the screening can be obtained as a recombinant protein using the nucleotide sequence of the marker gene. Based on the information of the marker gene, one skilled in the art can select any biological activity of the protein as an index for screening and a
20 measurement method based on the selected biological activity.

Alternatively, the screening method of the present invention may comprise the following steps:

- a) contacting a candidate compound with a cell into which a vector comprising the
transcriptional regulatory region of one or more marker genes and a reporter gene that
25 is expressed under the control of the transcriptional regulatory region has been
introduced, wherein the one or more marker genes are selected from the group
consisting of CML 1-296
- b) measuring the activity of said reporter gene; and
- c) selecting a compound that reduces the expression level of said reporter gene when said
30 marker gene is an up-regulated marker gene selected from the group consisting of
CML 1-190 or that enhances the expression level of said reporter gene when said

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marker gene is a down-regulated marker gene selected from the group consisting of CML 191-296, as compared to a control.

Suitable reporter genes and host cells are well known in the art. The reporter construct required for the screening can be prepared by using the transcriptional regulatory region of a marker gene. When the transcriptional regulatory region of a marker gene has been known to those skilled in the art, a reporter construct can be prepared by using the previous sequence information. When the transcriptional regulatory region of a marker gene remains unidentified, a nucleotide segment containing the transcriptional regulatory region can be isolated from a genome library based on the nucleotide sequence information of the marker gene.

The compound isolated by the screening is a candidate for drugs that inhibit the activity of the protein encoded by marker genes and can be applied to the treatment or prevention of CML.

Moreover, compound in which a part of the structure of the compound inhibiting the activity of proteins encoded by marker genes is converted by addition, deletion and/or replacement are also included in the compounds obtainable by the screening method of the present invention.

When administering the compound isolated by the method of the invention as a pharmaceutical for humans and other mammals, such as mice, rats, guinea-pigs, rabbits, chicken, cats, dogs, sheep, pigs, cattle, monkeys, baboons, and chimpanzees, the isolated compound can be directly administered or can be formulated into a dosage form using known pharmaceutical preparation methods. For example, according to the need, the drugs can be taken orally, as sugar-coated tablets, capsules, elixirs and microcapsules, or non-orally, in the form of injections of sterile solutions or suspensions with water or any other pharmaceutically acceptable liquid. For example, the compounds can be mixed with pharmaceutically acceptable carriers or media, specifically, sterilized water, physiological saline, plant-oils, emulsifiers, suspending agents, surfactants, stabilizers, flavoring agents, excipients, vehicles, preservatives, binders, and such, in a unit dose form required for generally accepted drug implementation. The amount of active ingredients in these preparations makes a suitable dosage within the indicated range acquirable.

Examples of additives that can be mixed to tablets and capsules are, binders such as gelatin, corn starch, tragacanth gum and arabic gum; excipients such as crystalline cellulose; swelling agents such as corn starch, gelatin and alginic acid; lubricants such as magnesium stearate; sweeteners such as sucrose, lactose or saccharin; and flavoring agents such as

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peppermint, Gaultheria adenoithrix oil and cherry. When the unit-dose form is a capsule, a liquid carrier, such as an oil, can also be further included in the above ingredients. Sterile composites for injections can be formulated following normal drug implementations using vehicles such as distilled water used for injections.

5 Physiological saline, glucose, and other isotonic liquids including adjuvants, such as D-sorbitol, D-mannnose, D-mannitol, and sodium chloride, can be used as aqueous solutions for injections. These can be used in conjunction with suitable solubilizers, such as alcohol, specifically ethanol, polyalcohols such as propylene glycol and polyethylene glycol, non-ionic surfactants, such as Polysorbate 80 (TM) and HCO-50.

10 Sesame oil or Soy-bean oil can be used as a oleaginous liquid and may be used in conjunction with benzyl benzoate or benzyl alcohol as a solubilizer and may be formulated with a buffer, such as phosphate buffer and sodium acetate buffer; a pain-killer, such as procaine hydrochloride; a stabilizer, such as benzyl alcohol and phenol; and an anti-oxidant. The prepared injection may be filled into a suitable ampule.

15 Methods well known to one skilled in the art may be used to administer the pharmaceutical composition of the present invention to patients, for example as intraarterial, intravenous, or percutaneous injections and also as intranasal, transbronchial, intramuscular or oral administrations. The dosage and method of administration vary according to the body-weight and age of a patient and the administration method; however, one skilled in the art can
20 routinely select a suitable method of administration. If said compound is encodable by a DNA, the DNA can be inserted into a vector for gene therapy and the vector administered to a patient to perform the therapy. The dosage and method of administration vary according to the body-weight, age, and symptoms of the patient but one skilled in the art can suitably select them.

 For example, although the dose of a compound that binds to the protein of the present
25 invention and regulates its activity depends on the symptoms, the dose is about 0.1 mg to about 100 mg per day, preferably about 1.0 mg to about 50 mg per day and more preferably about 1.0 mg to about 20 mg per day, when administered orally to a normal adult (weight 60 kg).

 When administering parenterally, in the form of an injection to a normal adult (weight 60 kg), although there are some differences according to the patient, target organ, symptoms and
30 method of administration, it is convenient to intravenously inject a dose of about 0.01 mg to about 30 mg per day, preferably about 0.1 to about 20 mg per day and more preferably about 0.1 to about 10 mg per day. Also, in the case of other animals too, it is possible to administer an

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amount converted to 60 kgs of body-weight.

Assessing the prognosis of a subject with CML

Also provided is a method of assessing the prognosis of a subject with CML by
5 comparing the expression of one or more CML sequences in a test cell population to the
expression of the sequences in a reference cell population derived from patients over a spectrum
of disease stages. By comparing gene expression of one or more CML sequences in the test cell
population and the reference cell population(s), or by comparing the pattern of gene expression
over time in test cell populations derived from the subject, the prognosis of the subject can be
10 assessed.

A decrease in expression of one or more of the sequences CML 191-296 compared to a
normal control or an increase of expression of one or more of the sequences CML 1-190
compared to a normal control indicates less favorable prognosis. An increase in expression of
one or more of the sequences CML 191-296 indicates a more favorable prognosis, and a
15 decrease in expression of sequences CML 1-190 indicates a more favorable prognosis for the
subject.

Kits

The invention also includes a CML-detection reagent, e.g., a nucleic acid that specifically
20 binds to or identifies one or more CML nucleic acids such as oligonucleotide sequences, which
are complementary to a portion of a CML nucleic acid or antibodies which bind to proteins
encoded by a CML nucleic acid. The reagents are packaged together in the form of a kit. The
reagents are packaged in separate containers, e.g., a nucleic acid or antibody (either bound to a
solid matrix or packaged separately with reagents for binding them to the matrix), a control
25 reagent (positive and/or negative), and/or a detectable label. Instructions (e.g., written, tape,
VCR, CD-ROM, etc.) for carrying out the assay are included in the kit. The assay format of the
kit is a Northern hybridization or a sandwich ELISA known in the art.

For example, CML detection reagent, is immobilized on a solid matrix such as a porous
strip to form at least one CML detection site. The measurement or detection region of the porous
30 strip may include a plurality of sites containing a nucleic acid. A test strip may also contain sites
for negative and/or positive controls. Alternatively, control sites are located on a separate strip
from the test strip. Optionally, the different detection sites may contain different amounts of

immobilized nucleic acids, *i.e.*, a higher amount in the first detection site and lesser amounts in subsequent sites. Upon the addition of test sample, the number of sites displaying a detectable signal provides a quantitative indication of the amount of CML present in the sample. The detection sites may be configured in any suitably detectable shape and are typically in the shape of a bar or dot spanning the width of a teststrip.

Alternatively, the kit contains a nucleic acid substrate array comprising one or more nucleic acid sequences. The nucleic acids on the array specifically identify one or more nucleic acid sequences represented by CML 1-296. The expression of 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 40 or 50 or more of the sequences represented by CML 1-296 are identified by virtue of the level of binding to an array test strip or chip. The substrate array can be on, *e.g.*, a solid substrate, *e.g.*, a "chip" as described in U.S. Patent No.5,744,305.

Arrays and pluralities

The invention also includes a nucleic acid substrate array comprising one or more nucleic acid sequences. The nucleic acids on the array specifically correspond to one or more nucleic acid sequences represented by CML 1-296. The level of expression of 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 40 or 50 or more of the sequences represented by CML 1-296 are identified by detecting nucleic acid binding to the array.

The invention also includes an isolated plurality (*i.e.*, a mixture of two or more nucleic acids) of nucleic acid sequences. The nucleic acid sequences are in a liquid phase or a solid phase, *e.g.*, immobilized on a solid support such as a nitrocellulose membrane. The plurality includes one or more of the nucleic acid sequences represented by CML 1-296. In various embodiments, the plurality includes 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 40 or 50 or more of the sequences represented by CML 1-296.

Methods of inhibiting CML

The invention provides a method for treating or alleviating a symptom of CML in a subject by decreasing expression or activity of CML 1-190 or increasing expression or activity of CML 191-296. Therapeutic compounds are administered prophylactically or therapeutically to a subject suffering from or at risk of (or susceptible to) developing CML. Such subjects are identified using standard clinical methods or by detecting an aberrant level of expression or activity of (*e.g.*, CML 1-296). Therapeutic agents include inhibitors of cell cycle regulation, cell

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proliferation, and protein kinase activity. Preferably, the inhibitor of kinase activity is not STI571. Alternatively, STI571 is administered together with one or more of the inhibitors of CML 1-296.

The therapeutic method includes increasing the expression, or function, or both of one or more gene products of genes whose expression is decreased ("underexpressed genes") in a CML cell relative to normal cells of the same cell type from which the CML cells are derived. In these methods, the subject is treated with an effective amount of a compound, which increases the amount of one or more of the underexpressed genes in the subject. Administration can be systemic or local. Therapeutic compounds include a polypeptide product of an underexpressed gene, or a biologically active fragment thereof, a nucleic acid encoding an underexpressed gene and having expression control elements permitting expression in the CML cells; for example an agent which increases the level of expression of such gene endogenous to the CML cells (i.e., which up-regulates expression of the underexpressed gene or genes). Administration of such compounds counter the effects of aberrantly-under expressed of the gene or genes in the subject's hematopoietic cells and improves the clinical condition of the subject.

The method also includes decreasing the expression, or function, or both, of one or more gene products of genes whose expression is aberrantly increased ("overexpressed gene") in hematopoietic cells including hematopoietic stem cells. Expression is inhibited in any of several ways known in the art. For example, expression is inhibited by administering to the subject a nucleic acid that inhibits, or antagonizes, the expression of the overexpressed gene or genes, e.g., an antisense oligonucleotide or small interfering RNA which disrupts expression of the overexpressed gene or genes.

As noted above, antisense nucleic acids corresponding to the nucleotide sequence of CML 1-190 can be used to reduce the expression level of the CML 1-190. Antisense nucleic acids corresponding to CML 1-190 that are up-regulated in CML are useful for the treatment of CML. Specifically, the antisense nucleic acids of the present invention may act by binding to the CML 1-190 or mRNAs corresponding thereto, thereby inhibiting the transcription or translation of the genes, promoting the degradation of the mRNAs, and/or inhibiting the expression of proteins encoded by the CML 1-190, finally inhibiting the function of the proteins. The term "antisense nucleic acids" as used herein encompasses both nucleotides that are entirely complementary to the target sequence and those having a mismatch of one or more nucleotides, so long as the antisense nucleic acids can specifically hybridize to the target sequences. For

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example, the antisense nucleic acids of the present invention include polynucleotides that have a homology of at least 70% or higher, preferably at 80% or higher, more preferably 90% or higher, even more preferably 95% or higher over a span of at least 15 continuous nucleotides.

Algorithms known in the art can be used to determine the homology.

5 The antisense nucleic acid derivatives of the present invention act on cells producing the proteins encoded by marker genes by binding to the DNAs or mRNAs encoding the proteins, inhibiting their transcription or translation, promoting the degradation of the mRNAs, and inhibiting the expression of the proteins, thereby resulting in the inhibition of the protein function.

10 An antisense nucleic acid derivative of the present invention can be made into an external preparation, such as a liniment or a poultice, by mixing with a suitable base material which is inactive against the derivative.

Also, as needed, the derivatives can be formulated into tablets, powders, granules, capsules, liposome capsules, injections, solutions, nose-drops and freeze-drying agents by adding
15 excipients, isotonic agents, solubilizers, stabilizers, preservatives, pain-killers, and such. These can be prepared by following known methods.

The antisense nucleic acids derivative is given to the patient by directly applying onto the ailing site or by injecting into a blood vessel so that it will reach the site of ailment. An antisense-mounting medium can also be used to increase durability and membrane-permeability.

20 Examples are, liposomes, poly-L-lysine, lipids, cholesterol, lipofectin or derivatives of these.

The dosage of the antisense nucleic acid derivative of the present invention can be adjusted suitably according to the patient's condition and used in desired amounts. For example, a dose range of 0.1 to 100 mg/kg, preferably 0.1 to 50 mg/kg can be administered.

25 The antisense nucleic acids of the invention inhibit the expression of the protein of the invention and is thereby useful for suppressing the biological activity of a protein of the invention. Also, expression-inhibitors, comprising the antisense nucleic acids of the invention, are useful since they can inhibit the biological activity of a protein of the invention.

The antisense nucleic acids of present invention include modified oligonucleotides. For example, thioated nucleotides may be used to confer nuclease resistance to an oligonucleotide.

30 Also, an siRNA against marker gene can be used to reduce the expression level of the marker gene. By the term "siRNA" is meant a double stranded RNA molecule which prevents translation of a target mRNA. Standard techniques of introducing siRNA into the cell are used,

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including those in which DNA is a template from which RNA is transcribed. In the context of the present invention, the siRNA comprises a sense nucleic acid sequence and an anti-sense nucleic acid sequence against an upregulated marker gene, such as CML 1-190. The siRNA is constructed such that a single transcript has both the sense and complementary antisense sequences from the target gene, e.g., a hairpin.

The method is used to alter the expression in a cell of an upregulated, e.g., as a result of malignant transformation of the cells. Binding of the siRNA to a transcript corresponding to one of the CML 1-190 in the target cell results in a reduction in the protein production by the cell. The length of the oligonucleotide is at least 10 nucleotides and may be as long as the naturally-occurring the transcript. Preferably, the oligonucleotide is 19-25 nucleotides in length. Most preferably, the oligonucleotide is less than 75, 50, 25 nucleotides in length.

The nucleotide sequence of the siRNAs were designed using an siRNA design computer program available from the Ambion website (http://www.ambion.com/techlib/misc/siRNA_finder.html). The computer program selects nucleotide sequences for siRNA synthesis based on the following protocol.

Selection of siRNA Target Sites:

1. Beginning with the AUG start codon of the object transcript, scan downstream for AA dinucleotide sequences. Record the occurrence of each AA and the 3' adjacent 19 nucleotides as potential siRNA target sites. Tuschl, et al. recommend against designing siRNA to the 5' and 3' untranslated regions (UTRs) and regions near the start codon (within 75 bases) as these may be richer in regulatory protein binding sites. UTR-binding proteins and/or translation initiation complexes may interfere with binding of the siRNA endonuclease complex.
2. Compare the potential target sites to the human genome database and eliminate from consideration any target sequences with significant homology to other coding sequences. The homology search can be performed using BLAST, which can be found on the NCBI server at: www.ncbi.nlm.nih.gov/BLAST/
3. Select qualifying target sequences for synthesis. At Ambion, preferably several target sequences can be selected along the length of the gene for evaluation.

The antisense oligonucleotide or siRNA of the invention inhibit the expression of the polypeptide of the invention and is thereby useful for suppressing the biological activity of the polypeptide of the invention. Also, expression-inhibitors, comprising the antisense

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oligonucleotide or siRNA of the invention, are useful in the point that they can inhibit the biological activity of the polypeptide of the invention. Therefore, a composition comprising the antisense oligonucleotide or siRNA of the present invention are useful in treating a CML.

Alternatively, function of one or more gene products of the overexpressed genes is inhibited by administering a compound that binds to or otherwise inhibits the function of the gene products. For example, the compound is an antibody which binds to the overexpressed gene product or gene products.

The present invention refers to the use of antibodies, particularly antibodies against a protein encoded by an up-regulated marker gene, or a fragment of the antibody. As used herein, the term "antibody" refers to an immunoglobulin molecule having a specific structure, that interacts (i.e., binds) only with the antigen that was used for synthesizing the antibody (i.e., the up-regulated marker gene product) or with an antigen closely related to it. Furthermore, an antibody may be a fragment of an antibody or a modified antibody, so long as it binds to one or more of the proteins encoded by the marker genes. For instance, the antibody fragment may be Fab, F(ab')₂, Fv, or single chain Fv (scFv), in which Fv fragments from H and L chains are ligated by an appropriate linker (Huston J. S. et al. Proc. Natl. Acad. Sci. U.S.A. 85:5879-5883 (1988)). More specifically, an antibody fragment may be generated by treating an antibody with an enzyme, such as papain or pepsin. Alternatively, a gene encoding the antibody fragment may be constructed, inserted into an expression vector, and expressed in an appropriate host cell (see, for example, Co M. S. et al. J. Immunol. 152:2968-2976 (1994); Better M. and Horwitz A. H. Methods Enzymol. 178:476-496 (1989); Pluckthun A. and Skerra A. Methods Enzymol. 178:497-515 (1989); Lamoyi E. Methods Enzymol. 121:652-663 (1986); Rousseaux J. et al. Methods Enzymol. 121:663-669 (1986); Bird R. E. and Walker B. W. Trends Biotechnol. 9:132-137 (1991)).

An antibody may be modified by conjugation with a variety of molecules, such as polyethylene glycol (PEG). The present invention provides such modified antibodies. The modified antibody can be obtained by chemically modifying an antibody. These modification methods are conventional in the field.

Alternatively, an antibody may be obtained as a chimeric antibody, between a variable region derived from a nonhuman antibody and a constant region derived from a human antibody, or as a humanized antibody, comprising the complementarity determining region (CDR) derived from a nonhuman antibody, the frame work region (FR) derived from a human antibody, and the

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constant region. Such antibodies can be prepared by using known technologies.

Cancer therapies directed at specific molecular alterations that occur in cancer cells have been validated through clinical development and regulatory approval of anti-cancer drugs such as trastuzumab (Herceptin) for the treatment of advanced breast cancer, imatinib methylate
5 (Gleevec) for chronic myeloid leukemia, gefitinib (Iressa) for non-small cell lung cancer (NSCLC), and rituximab (anti-CD20 mAb) for B-cell lymphoma and mantle cell lymphoma (Ciardiello F, Tortora G. A novel approach in the treatment of cancer: targeting the epidermal growth factor receptor. Clin Cancer Res. 2001 Oct;7(10):2958-70. Review.; Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, Fleming T, Eiermann W, Wolter J,
10 Pegram M, Baselga J, Norton L. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. N Engl J Med. 2001 Mar 15;344(11):783-92.; Rehwald U, Schulz H, Reiser M, Sieber M, Staak JO, Morschhauser F, Driessen C, Rudiger T, Muller-Hermelink K, Diehl V, Engert A. Treatment of relapsed CD20+ Hodgkin lymphoma with the monoclonal antibody rituximab is effective and well tolerated: results of a phase 2 trial
15 of the German Hodgkin Lymphoma Study Group. Blood. 2003 Jan 15;101(2):420-424.; Fang G, Kim CN, Perkins CL, Ramadevi N, Winton E, Wittmann S and Bhalla KN. (2000). Blood, 96, 2246-2253.). These drugs are clinically effective and better tolerated than traditional anti-cancer agents because they target only transformed cells. Hence, such drugs not only improve survival and quality of life for cancer patients, but also validate the concept of molecularly targeted
20 cancer therapy. Furthermore, targeted drugs can enhance the efficacy of standard chemotherapy when used in combination with it (Gianni L. (2002). Oncology, 63 Suppl 1, 47-56.; Klejman A, Rushen L, Morrione A, Slupianek A and Skorski T. (2002). Oncogene, 21, 5868-5876.). Therefore, future cancer treatments will probably involve combining conventional drugs with target-specific agents aimed at different characteristics of tumor cells such as angiogenesis and
25 invasiveness.

These modulatory methods are performed *ex vivo* or *in vitro* (e.g., by culturing the cell with the agent) or, alternatively, *in vivo* (e.g., by administering the agent to a subject). The method involves administering a protein or combination of proteins or a nucleic acid molecule or combination of nucleic acid molecules as therapy to counteract aberrant expression or activity of
30 the differentially expressed genes.

Diseases and disorders that are characterized by increased (relative to a subject not suffering from the disease or disorder) levels or biological activity of the genes may be treated

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with therapeutics that antagonize (*i.e.*, reduce or inhibit) activity of the overexpressed gene or genes. Therapeutics that antagonize activity are administered therapeutically or prophylactically.

Therapeutics that may be utilized include, *e.g.*, (i) a polypeptide, or analogs, derivatives, fragments or homologs thereof of the overexpressed or underexpressed sequence or sequences; (ii) antibodies to the overexpressed or underexpressed sequence or sequences; (iii) nucleic acids encoding the over or underexpressed sequence or sequences; (iv) antisense nucleic acids or nucleic acids that are "dysfunctional" (*i.e.*, due to a heterologous insertion within the coding sequences of coding sequences of one or more overexpressed or underexpressed sequences); (v) small interfering RNA (siRNA); or (vi) modulators (*i.e.*, inhibitors, agonists and antagonists that alter the interaction between an over/underexpressed polypeptide and its binding partner. The dysfunctional antisense molecules are utilized to "knockout" endogenous function of a polypeptide by homologous recombination (see, *e.g.*, Capecchi, Science 244: 1288-1292 1989). The term "siRNA" refers to a double stranded RNA molecule which prevents translation of a target mRNA. Standard techniques are used for introducing siRNA into cells, including those wherein DNA is used as the template to transcribe RNA. The siRNA comprises a sense nucleic acid sequence and an anti-sense nucleic acid sequence of the any one of the CML 1-190 gene. The siRNA is constructed such that a single transcript (double stranded RNA) has both the sense and complementary antisense sequences from the target gene, *e.g.*, a hairpin.

Diseases and disorders that are characterized by decreased (relative to a subject not suffering from the disease or disorder) levels or biological activity may be treated with therapeutics that increase (*i.e.*, are agonists to) activity. Therapeutics that upregulate activity may be administered in a therapeutic or prophylactic manner. Therapeutics that may be utilized include, but are not limited to, a polypeptide (or analogs, derivatives, fragments or homologs thereof) or an agonist that increases bioavailability.

Increased or decreased levels can be readily detected by quantifying peptide and/or RNA, by obtaining a patient cell sample and assaying it *in vitro* for RNA or peptide levels, structure and/or activity of the expressed peptides (or mRNAs of a gene whose expression is altered). Methods that are well-known within the art include, but are not limited to, immunoassays (*e.g.*, by Western blot analysis, immunoprecipitation followed by sodium dodecyl sulfate (SDS) polyacrylamide gel electrophoresis, immunocytochemistry, etc.) and/or hybridization assays to detect expression of mRNAs (*e.g.*, Northern assays, dot blots, *in situ* hybridization, etc.).

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Prophylactic administration occurs prior to the manifestation of overt clinical symptoms of disease, such that a disease or disorder is prevented or, alternatively, delayed in its progression.

Therapeutic methods includes contacting a cell with an agent that modulates one or more of the activities of the gene products of the differentially expressed genes. An agent that
5 modulates protein activity includes a nucleic acid or a protein, a naturally-occurring cognate ligand of these proteins, a peptide, a peptidomimetic, or other small molecule. For example, the agent stimulates one or more protein activities of one or more of a differentially under-expressed gene.

The present invention also relates to a method of treating or preventing CML in a
10 subject comprising administering to said subject a vaccine comprising a polypeptide encoded by a nucleic acid selected from the group consisting of CML 1-190 or an immunologically active fragment of said polypeptide, or a polynucleotide encoding the polypeptide or the fragment thereof. An administration of the polypeptide induce an anti-tumor immunity in a subject. To inducing anti-tumor immunity, a polypeptide encoded by a nucleic acid selected from the group
15 consisting of CML 1-190 or an immunologically active fragment of said polypeptide, or a polynucleotide encoding the polypeptide is administered. The polypeptide or the immunologically active fragments thereof are useful as vaccines against CML. In some cases the proteins or fragments thereof may be administered in a form bound to the T cell receptor (TCR) or presented by an antigen presenting cell (APC), such as macrophage, dendritic cell
20 (DC), or B-cells. Due to the strong antigen presenting ability of DC, the use of DC is most preferable among the APCs.

In the present invention, vaccine against CML refers to a substance that has the function to induce anti-tumor immunity upon inoculation into animals. According to the present invention, polypeptides encoded by CML 1-190 or fragments thereof were suggested to be HLA-
25 A24 or HLA-A*0201 restricted epitopes peptides that may induce potent and specific immune response against CML cells expressing CML 1-190. Thus, the present invention also encompasses method of inducing anti-tumor immunity using the polypeptides. In general, anti-tumor immunity includes immune responses such as follows:

- induction of cytotoxic lymphocytes against tumors,
- 30 - induction of antibodies that recognize tumors, and
- induction of anti-tumor cytokine production.

Therefore, when a certain protein induces any one of these immune responses upon

inoculation into an animal, the protein is decided to have anti-tumor immunity inducing effect. The induction of the anti-tumor immunity by a protein can be detected by observing in vivo or in vitro the response of the immune system in the host against the protein.

For example, a method for detecting the induction of cytotoxic T lymphocytes is well known. A foreign substance that enters the living body is presented to T cells and B cells by the action of antigen presenting cells (APCs). T cells that respond to the antigen presented by APC in antigen specific manner differentiate into cytotoxic T cells (or cytotoxic T lymphocytes; CTLs) due to stimulation by the antigen, and then proliferate (this is referred to as activation of T cells). Therefore, CTL induction by a certain peptide can be evaluated by presenting the peptide to T cell by APC, and detecting the induction of CTL. Furthermore, APC has the effect of activating CD4+ T cells, CD8+ T cells, macrophages, eosinophils, and NK cells. Since CD4+ T cells and CD8+ T cells are also important in anti-tumor immunity, the anti-tumor immunity inducing action of the peptide can be evaluated using the activation effect of these cells as indicators.

A method for evaluating the inducing action of CTL using dendritic cells (DCs) as APC is well known in the art. DC is a representative APC having the strongest CTL inducing action among APCs. In this method, the test polypeptide is initially contacted with DC, and then this DC is contacted with T cells. Detection of T cells having cytotoxic effects against the cells of interest after the contact with DC shows that the test polypeptide has an activity of inducing the cytotoxic T cells. Activity of CTL against tumors can be detected, for example, using the lysis of ⁵¹Cr-labeled tumor cells as the indicator. Alternatively, the method of evaluating the degree of tumor cell damage using ³H-thymidine uptake activity or LDH (lactose dehydrogenase)-release as the indicator is also well known.

Apart from DC, peripheral blood mononuclear cells (PBMCs) may also be used as the APC. The induction of CTL is reported that it can be enhanced by culturing PBMC in the presence of GM-CSF and IL-4. Similarly, CTL has been shown to be induced by culturing PBMC in the presence of keyhole limpet hemocyanin (KLH) and IL-7.

The test polypeptides confirmed to possess CTL inducing activity by these methods are polypeptides having DC activation effect and subsequent CTL inducing activity. Therefore, polypeptides that induce CTL against tumor cells are useful as vaccines against tumors. Furthermore, APC that acquired the ability to induce CTL against tumors by contacting with the polypeptides are useful as vaccines against tumors. Furthermore, CTL that acquired cytotoxicity

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due to presentation of the polypeptide antigens by APC can be also used as vaccines against tumors. Such therapeutic methods for tumors using anti-tumor immunity due to APC and CTL are referred to as cellular immunotherapy.

Generally, when using a polypeptide for cellular immunotherapy, efficiency of the CTL-induction is known to increase by combining a plurality of polypeptides having different structures and contacting them with DC. Therefore, when stimulating DC with protein fragments, it is advantageous to use a mixture of multiple types of fragments.

Alternatively, the induction of anti-tumor immunity by a polypeptide can be confirmed by observing the induction of antibody production against tumors. For example, when antibodies against a polypeptide are induced in a laboratory animal immunized with the polypeptide, and when growth of tumor cells is suppressed by those antibodies, the polypeptide can be determined to have an ability to induce anti-tumor immunity.

Anti-tumor immunity is induced by administering the vaccine of this invention, and the induction of anti-tumor immunity enables treatment and prevention of CML. Therapy against cancer or prevention of the onset of cancer includes any of the steps, such as inhibition of the growth of cancerous cells, involution of cancer, and suppression of occurrence of cancer. Decrease in mortality of individuals having cancer, decrease of tumor markers in the blood, alleviation of detectable symptoms accompanying cancer, and such are also included in the therapy or prevention of cancer. Such therapeutic and preventive effects are preferably statistically significant. For example, in observation, at a significance level of 5% or less, wherein the therapeutic or preventive effect of a vaccine against cell proliferative diseases is compared to a control without vaccine administration. For example, Student's t-test, the Mann-Whitney U-test, or ANOVA may be used for statistical analyses.

The above-mentioned protein having immunological activity or a vector encoding the protein may be combined with an adjuvant. An adjuvant refers to a compound that enhances the immune response against the protein when administered together (or successively) with the protein having immunological activity. Examples of adjuvants include cholera toxin, salmonella toxin, alum, and such, but are not limited thereto. Furthermore, the vaccine of this invention may be combined appropriately with a pharmaceutically acceptable carrier. Examples of such carriers are sterilized water, physiological saline, phosphate buffer, culture fluid, and such. Furthermore, the vaccine may contain as necessary, stabilizers, suspensions, preservatives, surfactants, and such. The vaccine is administered systemically or locally. Vaccine

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administration may be performed by single administration, or boosted by multiple administrations.

When using APC or CTL as the vaccine of this invention, tumors can be treated or prevented, for example, by the ex vivo method. More specifically, PBMCs of the subject receiving treatment or prevention are collected, the cells are contacted with the polypeptide ex vivo, and following the induction of APC or CTL, the cells may be administered to the subject. APC can be also induced by introducing a vector encoding the polypeptide into PBMCs ex vivo. APC or CTL induced in vitro can be cloned prior to administration. By cloning and growing cells having high activity of damaging target cells, cellular immunotherapy can be performed more effectively. Furthermore, APC and CTL isolated in this manner may be used for cellular immunotherapy not only against individuals from whom the cells are derived, but also against similar types of tumors from other individuals.

Furthermore, a pharmaceutical composition for treating or preventing a cell proliferative disease, such as cancer, comprising a pharmaceutically effective amount of the polypeptide of the present invention is provided. The pharmaceutical composition may be used for raising anti tumor immunity.

Pharmaceutical compositions for inhibiting CML

Pharmaceutical formulations include those suitable for oral, rectal, nasal, topical (including buccal and sub-lingual), vaginal or parenteral (including intramuscular, sub-cutaneous and intravenous) administration, or for administration by inhalation or insufflation. Preferably, administration is intravenous. The formulations are optionally packaged in discrete dosage units

Pharmaceutical formulations suitable for oral administration include capsules, cachets or tablets, each containing a predetermined amount of the active ingredient. Formulations also include powders, granules or solutions, suspensions or emulsions. The active ingredient is optionally administered as a bolus electuary or paste. Tablets and capsules for oral administration may contain conventional excipients such as binding agents, fillers, lubricants, disintegrant or wetting agents. A tablet may be made by compression or molding, optionally with one or more formulation ingredients. Compressed tablets may be prepared by compressing in a suitable machine the active ingredients in a free-flowing form such as a powder or granules, optionally mixed with a binder, lubricant, inert diluent, lubricating, surface active or dispersing agent. Molded tablets may be made by molding in a suitable machine a mixture of the powdered

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compound moistened with an inert liquid diluent. The tablets may be coated according to methods well known in the art. Oral fluid preparations may be in the form of, for example, aqueous or oily suspensions, solutions, emulsions, syrups or elixirs, or may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may contain conventional additives such as suspending agents, emulsifying agents, non-aqueous vehicles (which may include edible oils), or preservatives. The tablets may optionally be formulated so as to provide slow or controlled release of the active ingredient therein. A package of tablets may contain one tablet to be taken on each of the month. The formulation or dose of medicament varies with respect to the phase (chronic, accelerated, or blast crisis) of the CML.

Formulations for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The formulations may be presented in unit dose or multi-dose containers, for example sealed ampoules and vials, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example, saline, water-for-injection, immediately prior to use. Alternatively, the formulations may be presented for continuous infusion. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules and tablets of the kind previously described.

Formulations for rectal administration include suppositories with standard carriers such as cocoa butter or polyethylene glycol. Formulations for topical administration in the mouth, for example buccally or sublingually, include lozenges, which contain the active ingredient in a flavored base such as sucrose and acacia or tragacanth, and pastilles comprising the active ingredient in a base such as gelatin and glycerin or sucrose and acacia. For intra-nasal administration the compounds of the invention may be used as a liquid spray or dispersible powder or in the form of drops. Drops may be formulated with an aqueous or non-aqueous base also comprising one or more dispersing agents, solubilizing agents or suspending agents.

For administration by inhalation the compounds are conveniently delivered from an insufflator, nebulizer, pressurized packs or other convenient means of delivering an aerosol spray. Pressurized packs may comprise a suitable propellant such as dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the

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case of a pressurized aerosol, the dosage unit may be determined by providing a valve to deliver a metered amount.

Alternatively, for administration by inhalation or insufflation, the compounds may take the form of a dry powder composition, for example a powder mix of the compound and a
5 suitable powder base such as lactose or starch. The powder composition may be presented in unit dosage form, in for example, capsules, cartridges, gelatin or blister packs from which the powder may be administered with the aid of an inhalator or insufflators.

Other formulations include implantable devices and adhesive patches; which release a therapeutic agent.

10 When desired, the above described formulations, adapted to give sustained release of the active ingredient, may be employed. The pharmaceutical compositions may also contain other active ingredients such as antimicrobial agents, immunosuppressants or preservatives.

It should be understood that in addition to the ingredients particularly mentioned above, the formulations of this invention may include other agents conventional in the art having regard
15 to the type of formulation in question, for example, those suitable for oral administration may include flavoring agents.

Preferred unit dosage formulations are those containing an effective dose, as recited below, or an appropriate fraction thereof, of the active ingredient.

For each of the aforementioned conditions, the compositions, e.g., polypeptides and
20 organic compounds are administered orally or via injection at a dose of from about 0.1 to about 250 mg/kg per day. The dose range for adult humans is generally from about 5 mg to about 17.5 g/day, preferably about 5 mg to about 10 g/day, and most preferably about 100 mg to about 3 g/day. Tablets or other unit dosage forms of presentation provided in discrete units may conveniently contain an amount which is effective at such dosage or as a multiple of the same,
25 for instance, units containing about 5 mg to about 500 mg, usually from about 100 mg to about 500 mg.

The dose employed will depend upon a number of factors, including the age and sex of the subject, the precise disorder being treated, and its severity. Also the route of administration may vary depending upon the condition and its severity.

30 The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims. The following examples illustrate the identification and characterization of genes differentially expressed in CML cells.

EXAMPLE 1: PREPARATION OF TEST SAMPLES

Samples obtained from diseased cells and normal cells, *e.g.*, mononuclear cells from peripheral blood, were evaluated to identify genes which are differently expressed in a disease state, *e.g.*, CML. The assays were carried out as follows.

Patients and samples

Peripheral blood samples were obtained from 27 CML patients prior to treatment with STI571. Each patient was then enrolled into a phase II study of STI571. To characterize CML cells, mRNA from 27 samples in which more than 65% of cells had been positive for the Ph chromosome prior to treatment by a FISH analysis detecting a *bcr/abl* fusion gene (13) were analyzed on a cDNA-microarray system. Of the 27, two cases were in accelerated phase and three cases were in blast crisis phase (Table 1). A mixture of mononuclear cells from peripheral blood from eleven healthy volunteers was used as a control.

Table 1 Clinicopathological features of patients examined

Patient's ID	Age (y)	Sex	Ph (+) (%) a	Phase
CML002	71	F	78	Chronic
CML003	66	M	69	Chronic
CML004	55	F	75.5	Chronic
CML008	61	F	75	Chronic
CML009	68	M	80.5	Chronic
CML010	56	M	65.5	Chronic
CML013	59	F	87	Chronic
CML014	47	M	83.5	Chronic
CML015	63	F	72	Chronic
CML018	57	M	83.5	Chronic
CML019	23	M	79	Chronic
CML021	57	M	79.5	Chronic
CML022	69	M	72.5	Chronic
CML023	68	F	76	Chronic
CML025	44	M	76	Chronic
CML027	35	M	75.5	Chronic
CML029	45	F	73	Chronic
CML030	61	M	65.5	Chronic

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CML033	56	M	66	Chronic
CML036	48	M	77	Chronic
CML047	32	F	85.5	Chronic
CML054	32	M	71	Chronic

RNA Preparation and T7-based RNA amplification

Mononuclear cells were prepared using Ficoll (Amersham Biosciences, Buckinghamshire, UK) and total RNA was extracted using TRIzol (Life Technologies, Inc., Grand Island, NY) according to the manufacturer's instructions. After treatment with DNase I (Nippon Gene, Tokyo, Japan), T7-based RNA amplification was carried out (14). Two rounds of amplification using 2 µg of total RNA as starting material yielded 40-100 µg of amplified RNA (aRNA). For control samples from healthy volunteers, two rounds of T7-based RNA amplification was also performed to obtain a sufficient amount of aRNA. RNA amplified by this method accurately reflects the proportions in the original RNA source, as confirmed by semi-quantitative reverse transcription-polymerase chain reaction (RT-PCR) experiments, in which data from the microarrays were consistent with results from RT-PCR whether total RNA or amplified aRNA was used as the template (14).

Preparation of the cDNA microarray and hybridization

A genome-wide cDNA microarray was fabricated with 23,040 cDNAs selected from the UniGene database (build #131) of the National Center for Biotechnology Information (NCBI). To obtain cDNAs for spotting on the glass slides, RT-PCR was performed for each gene as described previously. (15) The PCR products were spotted on type 7 glass slides (Amersham Biosciences) by Microarray Spotter Generation III (Amersham Biosciences); 4,608 genes were spotted in duplicate on a single slide. Five different sets of slides (total 23,040 genes) were prepared, on each of which the same 52 housekeeping genes and two negative-control genes were spotted as well. Labeling, hybridization, washing, scanning, and quantification of signals were performed as described previously (14) except that all processes were carried out with an Automated Slide Processor (15).

Quantification of signals and data analysis

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The intensity of each hybridization signal was calculated photometrically using the ArrayVision computer program (Amersham Biosciences). Each slide contained 52 housekeeping genes, and the Cy5/Cy3 ratio for each gene's expression was adjusted so that the averaged Cy5/Cy3 ratio of the panel of housekeeping genes was 1.0. A cut-off value was assigned to each microarray slide, using variance analysis. If both Cy3 and Cy5 signal intensities were lower than the cut-off values, the expression level of the corresponding gene in that sample was assessed as absent. For other genes the Cy5/Cy3 ratio was calculated using raw data of each sample.

10 EXAMPLE 2: IDENTIFICATION OF CML – ASSOCIATED GENES

The relative expression ratio of each gene (Cy5/Cy3 intensity ratio) was classified into one of four categories: (1) highly up-regulated (expression ratio more than 5.0 in more than 50% of the informative cases); (2) highly down-regulated (expression ratio less than 0.2 in more than 50% of the informative cases); (3) low expression (expression ratio between 0.2 and 5.0 in more than 50% of the informative cases); and (4) not expressed (or slight expression but under the cut-off level for detection). These categories were used to detect a set of genes whose changes in expression ratios were common among samples as well as specific to a certain subgroup. To detect candidate genes that were commonly up- or down-regulated in CML cells, the overall expression patterns of 23,040 genes were screened to select genes with expression ratios of more than 5.0 or less than 0.2 that were present in more than 50% of chronic phase of the CML cases categorized as (1), (2), or (3).

Identification of genes with clinically relevant expression patterns in CML cells

The expression patterns of approximately 23,000 genes in CML cells were examined using cDNA microarray. Individual data was excluded when both Cy5 and Cy3 signals were under cut-off values. The computational analysis identified commonly highly up-regulated or down-regulated genes in CML cells; 190 genes revealed the expression ratio of >5.0 in more than 50% of informative cases and 106 genes showed the expression ratio of <0.2 in more than 50% of informative cases as down-regulated genes.

One hundred ninety genes were found to be highly up-regulated. The upregulated genes included genes encoding proteins involved in cell cycle regulation, growth promotion, and

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transcriptional activation and those having protein kinase activity. Many of them were shown to be over-expressed in other carcinomas. For example, MYB, a transcriptional activator that causes acute leukemia and transforms only hematopoietic cells (16), was highly expressed in over 90 % of the chronic phase of CML cells. GATA-binding protein 2 (GATA2), also a transcriptional activator which regulates endotherin-1 gene expression in endothelial cells (17), was reported to be activated in 93% of acute myeloid leukemia (AML), 70% of acute lymphoblastic leukemia (ALL), and 83% of CML (18). In particular, 28 genes for example, ribonuclease RNase A family 3, (RNASE3), bactericidal/permeability-increasing protein (BP1), defensin alpha 1, myeloid-related sequence (DEFA1), aminolevulinate, delta-synthase 1 (ALAS1), elastase 2, neutrophil (ELA2), cathepsin G (CTSG), matrix metalloprotease 9 (MMP-9), haptoglobin-related protein (HPR), urokinase plasminogen activator, (UPLA), haptoglobin (HP), H3 histone family, member J (H3FJ), and hemoglobin, zeta (HBZ) were overexpressed in all of the informative samples in this study (*see* Table 3). MMP-9, an enzyme to degrade collagen type IV, is thought to be associated with the transmigration and degradation of the extracellular matrix structures of tissue and blood vessels. The expression of MMP-9 was enhanced in mononuclear cells of CML patients (19). Moreover, primary human Ph⁺ cells were reported to secrete various angiogenesis factors including MMP-9 (20). Thus, overexpression of MMP-9 might play an important role in the pathogenesis of CML. Furthermore, members in the hemoglobin family, for example, zeta (HBZ), beta (HBB), gamma G (HBG2), delta (HBD), and alpha 2 (HBA2) were overexpressed in more than 80% of the informative cases. In addition, haptoglobin (HP) and haptoglobin-related protein (HPR) also showed enhanced expression in all informative cases. Recent studies have shown that bcr/abl expression induced hemoglobin (Hb) production in HL-60/BCR-Abl cells or CML cells (21), (22). This suggests that the constitutively activated tyrosine kinase bcr-abl enhanced survival and expansion of hematopoietic progenitor cells.

One hundred six genes were found to be significantly down-regulated in the chronic phase of CML (*see* Table 4). The genes of known function included the SH3-domain GRB2-line 2 (SH3GL2), PCAF associated factor 65 beta (PAF65B), heparan sulfate 6-O-sulfotransferase (HS6ST), immunoglobulin heavy constant gamma 3 (IGHG3), heat shock 27kD protein 2 (HSPB2), and prostaglandin D synthase gene (PTGDS) genes whose expression was suppressed in more than 90% of informative cases. A number of transcriptional negative regulators like DNA-dependent protein kinase catalytic subunit-interacting protein 2 (KIP2) were also included.

KIP2 is a negative regulator of cell proliferation and arrests cells at the G1 phase. KIP2 was down-regulated in approximately 60% of the informative CML cases. Therefore, its down-regulation may confer continuous proliferative properties of leukemic cells.

Some of the significantly down-regulated genes reflect the difference in the cell types, the lymphocyte-specific genes such as genes encoding immune components, *e.g.*, immunoglobulins, and complement component 2 (C2) as well as the markers for lymphocytes such as CD7, CD3E, CD79A, CD3Z, CD6, CD4, and CD79B antigens, interferon regulatory factor 4 (IRF4), and interleukin 7 receptor (IR7R) (*see* Table 4). The majority of the cells contained in the chronic phase of CML cells used in this study corresponded to blast cells of myeloid lineage. Although the peripheral white cell used as universal controls contained cells of both myeloid and lymphoid lineage, lymphocytes accounted for only a small population of the cells in the sample. This might represent that the population of lymphocyte in CML patients decreased compared with blood of healthy individuals.

TABLE3 UP-REGULATED GENES

CML As signment	ratio(%) 1)	informati ve cases	ratio>5 2)	Accession No.	Symbol	Gene name
1	100	22	22	X16545	RNASE3	ribonuclease, RNase A family, 3 (eosinophil cationic protein)
2	100	22	22	J04739	BPI	bactericidal/permeability-increasing protein
3	100	22	22	M21130	DEFA1	defensin, alpha 1, myeloid-related sequence
4	100	22	22	X56351	ALAS1	aminolevulinate, delta-, synthase 1
5	100	21	21	M16117	CTSG	cathepsin G
6	100	21	21	M34379	ELA2	elastase 2, neutrophil
7	100	20	20	J05070	MMP9	matrix metalloproteinase 9 (gelatinase B, 92kD gelatinase, 92kD type IV collagenase)
8	100	20	20	K03431	HPR	haptoglobin-related protein
9	100	20	20	F21002		ESTs
10	100	19	19	X17042	PRG1	proteoglycan 1, secretory granule
11	100	19	19	M14502	ARG1	arginase, liver
12	100	18	18	L06895	MAD	MAX dimerization protein
13	100	18	18	M69199	G0S2	putative lymphocyte G0/G1 switch gene
14	100	17	17	K01763	HP	haptoglobin

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15	100	16	16	H23213		ESTs
16	100	16	16	Z98744	H3FJ	H3 histone family, member J
17	100	15	15	M24173	HBZ	hemoglobin, zeta
18	100	15	15	X02419	PLAU	plasminogen activator, urokinase
19	100	15	15	H48537		ESTs
20	100	15	15	AA191449	KIAA1254	KIAA1254 protein
21	100	15	15	AI022380		ESTs
22	100	15	15	T03595		Homo sapiens cDNA FLJ12688 fis, clone NT2RM4002534
23	100	14	14	X57129	H1F2	H1 histone family, member 2
24	100	14	14	AA382504		ESTs
25	100	13	13	R26792	GCL	grancalcin
26	100	12	12	AA815247		EST
27	100	11	11	M81637	GCL	grancalcin
28	100	11	11	AA446449		EST
29	95.45	22	21	M83202	LTF	lactotransferrin
30	95.45	22	21	V00497	HBB	hemoglobin, beta
31	95.45	22	21	U01317	HBD	hemoglobin, delta
32	95.24	21	20	AA489915	HBG2	hemoglobin, gamma G
33	95	20	19	AI023753		ESTs
34	94.74	19	18	X83006	LCN2	lipocalin 2 (oncogene 24p3)
35	93.75	16	15	AI040591		ESTs
36	93.33	15	14	AA855085	NCOA4	nuclear receptor coactivator 4
37	91.67	12	11	M65085	FSHR	follicle stimulating hormone receptor
38	91.67	12	11	AA398536		ESTs
39	91.67	12	11	AA843554		ESTs
40	90.91	22	20	S81914	IER3	immediate early response 3
41	90.91	22	20	M27717	CPA3	carboxypeptidase A3 (mast cell)
42	90.91	22	20	U22376	MYB	v-myb avian myeloblastosis viral oncogene homolog
43	90.91	11	10	U32315	STX3A	syntaxin 3A
44	90.91	11	10	AA774546	NXF3	nuclear RNA export factor 3
45	90.48	21	19	AI015633		Solute carrier family 26, member 8
46	89.47	19	17	M33987	CA1	carbonic anhydrase I
47	88.89	18	16	AA004412		ESTs
48	88.89	18	16	AI056326		ESTs
49	88.24	17	15	AA825819	LOC55871	COBW-like protein
50	86.67	15	13	X65614	S100P	S100 calcium-binding protein P

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51	86.67	15	13	L27711	CDKN3	cyclin-dependent kinase inhibitor 3 (CDK2-associated dual specificity phosphatase)
52	86.67	15	13	AA418448		ESTs
53	86.67	15	13	AA235222	LOC51053	geminin
54	86.36	22	19	M62831	ETR101	immediate early protein
55	86.36	22	19	L01664	CLC	Charot-Leyden crystal protein
56	86.36	22	19	V00493	HBA2	hemoglobin, alpha 2
57	86.36	22	19	X00637	HP	haptoglobin
58	86.36	22	19	W63676		ESTs
59	85.71	21	18	U01317	HBD	hemoglobin, delta
60	85.71	14	12	N52485	DKFZP434	DKFZP434O125 protein O125
61	85.71	14	12	AI335266	FER1L3	fer (C.elegans)-like 3 (myoferlin)
62	85	20	17	Z83821	ALAS2	aminolevulinate, delta-, synthase 2 (sideroblastic/hypochromic anemia)
63	85	20	17	AA318275	FTH1	Ferritin, heavy polypeptide 1
64	84.62	13	11	M16827	ACADM	acyl-Coenzyme A dehydrogenase, C-4 to C2 straight chain
65	83.33	12	10	AA355657	CTSG	cathepsin G
66	81.82	22	18	D14874	ADM	adrenomedullin
67	81.82	22	18	M69043	NFKBIA	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha
68	81.82	22	18	X55656	HBG2	hemoglobin, gamma G
69	81.82	22	18	X68277	DUSP1	dual specificity phosphatase 1
70	81.82	22	18	W76477	JUN	v-jun avian sarcoma virus 17 oncogene homolog
71	81.82	22	18	X56351	ALAS1	aminolevulinate, delta-, synthase 1
72	81.82	11	9	AI168658	FECH	ferrochelatase (protoporphyrinogen oxidase)
73	81.82	11	9	AA256671	FLJ21939	hypothetical protein FLJ21939 similar to 5-azacytidine induced gene 2
74	80	15	12	AI819724	COL1A1	collagen, type I, alpha 1
75	78.95	19	15	M96839	PRTN3	proteinase 3 (serine proteinase, neutrophil, Wegener granulomatosis autoantigen)
76	78.95	19	15	R78436	GATA2	GATA-binding protein 2
77	78.57	14	11	AA449950	KIAA1016	KIAA1016 protein
78	77.78	18	14	AA456242	FSP-2	fibrousheathin II
79	77.78	18	14	AI357641	CDKN2C	cyclin-dependent kinase inhibitor 2C

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						tor 2C (p18, inhibits CDK4)
80	77.78	18	14	M13692	ORM1	orosomucoid 1
81	77.78	18	14	D90145	SCYA3L1	small inducible cytokine A3-like 1
82	77.78	18	14	AA903016	HM74	putative chemokine receptor; GTP-binding protein
83	77.27	22	17	AI128538	LOC51312	mitochondrial solute carrier
84	76.92	13	10	AF041245	HCRTR2	hypocretin (orexin) receptor 2
85	76.47	17	13	D13752	CYP11B2	cytochrome P450, subfamily XIB (steroid 11-beta-hydroxylase), polypeptide 2
86	76.47	17	13	AI360412	ALS2CR2	Amyotrophic lateral sclerosis 2 (juvenile) chromosome region, candidate 2
87	76.47	17	13	N30414		ESTs
88	76.19	21	16	AA126620	C8FW	Phosphoprotein regulated by mitogenic pathways
89	75	20	15	M33492	TPSB1	tryptase beta 1
90	75	20	15	AF043584	IGL	immunoglobulin lambda chain
91	75	16	12	AI000650		ESTs
92	75	16	12	AW337343	PTP4A1	protein tyrosine phosphatase type IVA, member 1
93	73.68	19	14	AA043835	DAPP1	dual adaptor of phosphotyrosine and 3-phosphoinositides
94	73.33	15	11	AA449227		EST
95	72.73	22	16	U21847	TIEG	TGFB inducible early growth response
96	72.73	11	8	U77942	STX7	syntaxin 7
97	72.73	11	8	AA345854	ITGA3	integrin, alpha 3 (antigen CD49C, alpha 3 subunit of VLA-3 receptor)
98	72.73	11	8	AA314457	LOC56994	cholinephosphotransferase 1
99	72.73	11	8	AI718618	BIRC2	baculoviral IAP repeat-containing 2
100	72.73	11	8	AI276054	FRAT2	Frequently rearranged in advanced T-cell lymphomas 2
101	72.22	18	13	D86724	ARG2	arginase, type II
102	72.22	18	13	X06233	S100A9	S100 calcium-binding protein A9 (calgranulin B)
103	72.22	18	13	AA973757	STX3A	syntaxin 3A
104	72.22	18	13	AF068754	HSBP1	heat shock factor binding protein 1
105	71.43	14	10	H97976		ESTs
106	70.59	17	12	D14657	KIAA0101	KIAA0101 gene product
107	70	20	14	AI056641	FLJ22833	hypothetical protein FLJ22833

108	70	20	14	AA677931		ESTs
109	69.23	13	9	AB003476	AKAP12	A kinase (PRKA) anchor prote in (gravin) 12
110	68.75	16	11	AA854469	RNF6	ring finger protein (C3H2C3 t ype) 6
111	68.18	22	15	AA327207		ESTs
112	66.67	21	14	AA772709		Homo sapiens cDNA FLJ1352 2 fis, clone PLACE1005884
113	66.67	21	14	U46254		ESTs
114	66.67	21	14	AI087002		ESTs
115	66.67	18	12	AW237266	ASAH	N-acylsphingosine amidohydr olase (acid ceramidase)
116	66.67	12	8	H53099	NDUFA10	NADH dehydrogenase (ubiqui none) 1 alpha subcomplex, 10 (42kD)
117	66.67	12	8	T03044		EST
118	65	20	13	W96110	ZNF281	zinc finger protein 281
119	64.71	17	11	AA629596	DKFZP564	DKFZP564D177 protein D177
120	64.29	14	9	M23161	THE1	Human transposon-like eleme nt mRNA
121	64.29	14	9	D20186	DKFZp762	hypothetical protein DKFZp76 O076 2O076
122	64.29	14	9	H80325	BAZ1A	bromodomain adjacent to zinc finger domain, 1A
123	63.64	22	14	U21847	TIEG	TGFB inducible early growth r esponse
124	63.64	11	7	M31452	C4BPA	complement component 4-bin ding protein, alpha
125	63.64	11	7	U39231	GIPR	gastric inhibitory polypeptide r eceptor
126	63.64	11	7	AI262031	ATP10D	ATPase, Class V, type 10D
127	63.64	11	7	AF022385	PDCD10	programmed cell death 10
128	63.64	11	7	N58488		EST
129	63.64	11	7	AI016419		ESTs
130	62.5	16	10	H71303	KIAA0481	KIAA0481 gene product
131	62.5	16	10	H06819	FLJ10846	hypothetical protein FLJ10846
132	62.5	16	10	W93000		ESTs
133	62.5	16	10	W37916	HCF-2	host cell factor 2
134	61.9	21	13	AF016833	MGAM	maltase-glucoamylase (alpha- glucosidase)
135	61.54	13	8	X00948	RLN2	relaxin 2 (H2)
136	61.54	13	8	U55206	GGH	gamma-glutamyl hydrolase (c onjugase, folylpolygammaglut amyl hydrolase)

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137	61.54	13	8	AA308062	S100P	S100 calcium-binding protein P
138	61.54	13	8	AA731746		ESTs
139	60	20	12	AI078178		ESTs
140	60	15	9	M95809	GTF2H1	general transcription factor II H, polypeptide 1 (62kD subunit)
141	60	15	9	AA401589		ESTs
142	59.09	22	13	M93056	SERPINB1	serine (or cysteine) proteinase inhibitor, clade B (ovalbumin), member 1
143	59.09	22	13	AA436509	IERS5	Immediate early response 5
144	58.82	17	10	L10101	SRY	sex determining region Y
145	58.33	12	7	W87690		Homo sapiens cDNA: FLJ23173 fis, clone LNG10019
146	57.89	19	11	AK025906		Homo sapiens cDNA: FLJ22253 fis, clone HRC02763
147	57.89	19	11	AA548765	SMARCB1	SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily b, member 1
148	57.14	21	12	AI014551		ESTs
149	57.14	21	12	AA313441		Homo sapiens cDNA FLJ11838 fis, clone HEMBA1006624, weakly similar to DNA/PANTOTHENATE METABOLISM FLAVOPROTEIN HOMOLOG
150	57.14	14	8	D49958	GPM6A	glycoprotein M6A
151	57.14	14	8	AA778025		ESTs
152	57.14	14	8	AA047169		Homo sapiens cDNA: FLJ22756 fis, clone KAIA0791
153	56.25	16	9	L16464	ETV3	ets variant gene 3
154	56.25	16	9	AF011468	STK15	serine/threonine kinase 15
155	56.25	16	9	AI188389	C11ORF15	chromosome 11 open reading frame 15
156	55	20	11	AA847136	CSF2RB	Colony stimulating factor 2 receptor, beta, low-affinity (granulocyte-macrophage)
157	55	20	11	AI025297	KLF7	Kruppel-like factor 7 (ubiquitous)
158	54.55	22	12	M60974	GADD45A	growth arrest and DNA-damage-inducible, alpha
159	54.55	22	12	X56351	ALAS1	aminolevulinate, delta-, synthase 1
160	54.55	11	6	U93869	RPC39	polymerase (RNA) III (DNA d

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						irected) (39kD)
161	54.55	11	6	AA551628	FLJ10260	hypothetical protein FLJ10260
162	54.55	11	6	AI268502		ESTs
163	54.55	11	6	AI768505	KIAA0707	KIAA0707 protein
164	54.55	11	6	W70293		ESTs
165	53.85	13	7	M28443	AMY2A	amylase, alpha 2A; pancreatic
166	53.85	13	7	L77571	DGS-A	DiGeorge syndrome gene A
167	53.85	13	7	AA101229		ESTs
168	53.33	15	8	X51699	BGLAP	bone gamma-carboxyglutamate (gla) protein (osteocalcin)
169	53.33	15	8	R86067		ESTs, Weakly similar to KIAA1353 protein [H.sapiens]
170	53.33	15	8	U57961	13CDNA73	putative gene product
171	53.33	15	8	AI025822		EST
172	53.33	15	8	AI150469		ESTs
173	52.94	17	9	M23204	OAT	ornithine aminotransferase (gyrate atrophy)
174	52.63	19	10	L04270	LTBR	lymphotoxin beta receptor (TNFR superfamily, member 3)
175	52.63	19	10	AW511361	SLC29A1	solute carrier family 29 (nucleoside transporters), member 1
176	52.63	19	10	AA027229		ESTs, Weakly similar to F45E12.5 [C.elegans]
177	52.63	19	10	AI201982		Homo sapiens cDNA FLJ11516 fis, clone HEMBA1002328
178	52.38	21	11	AA706319		ESTs
179	52.38	21	11	W05688		ESTs
180	50	20	10	X56741	MEL	mel transforming oncogene (derived from cell line NK14)- RAB8 homolog
181	50	18	9	AI032402		ESTs
182	50	16	8	F10728		ESTs
183	50	16	8	AA576399		ESTs
184	50	16	8	T36260	SEC23B	Sec23 (S. cerevisiae) homolog B
185	50	16	8	AJ250075	CH1	membrane protein CH1
186	50	12	6	AI187066		ESTs
187	50	12	6	AA586974	PI3	protease inhibitor 3, skin-derived (SKALP)
188	50	14	7	AA639599	SLC12A2	solute carrier family 12 (sodium/potassium/chloride transporters), member 2
189	50	14	7	AA807551		ESTs
190	50	14	7	AA503224		ESTs

Accession numbers and gene symbols were retrieved from the Unigene Databases (build#131).

TABLE4 DOWN-REGULATED GENES

CML Assignment	ratio(%) 1)	informati ve cases	ratio<5 2)	Accession No.	Symbol	Gene name
191	100	16	16	AF036268	SH3GL2	SH3-domain GRB2-like 2
192	94.74	19	18	AF069736	PAF65B	PCAF associated factor 65 beta
193	93.75	16	15	AB006179	HS6ST	heparan sulfate 6-O-sulfotransferase
194	93.33	15	14	AA806043	IGHG3	immunoglobulin heavy constant gamma 3 (G3m marker)
195	90.91	11	10	D89617	HSPB2	heat shock 27kD protein 2
196	90.48	21	19	M61900	PTGDS	prostaglandin D synthase gene
197	88.24	17	15	M73780	ITGB8	integrin, beta 8
198	88.24	17	15	M60450	KCNA4	potassium voltage-gated channel, shaker-related subfamily, member 4
199	87.5	16	14	U28369	SEMA3B	sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3B
200	86.67	15	13	M13149	HRG	histidine-rich glycoprotein
201	86.67	15	13	AI089023	FXYP7	FXYP domain-containing ion transport regulator 7
202	86.36	22	19	D00749	CD7	CD7 antigen
203	85	20	17	M88468	MVK	mevalonate kinase (mevalonic aciduria)
204	84.62	13	11	AF040723	HAP1	huntingtin-associated protein 1 (neuroan 1)
205	84.21	19	16	M24405	TCF3	transcription factor 3 (E2A immunoglobulin enhancer binding factors E12/E47)
206	82.35	17	14	M81886	GRIA1	glutamate receptor, ionotropic, AMPA 1
207	82.35	17	14	M25809	ATP6B1	ATPase, H ⁺ transporting, lysosomal (vacuolar proton pump), beta polypeptide, 56/58kD, isoform 1
208	81.82	22	18	R72651		ESTs, Weakly similar to PLK_HUMAN PROTEOGLYCAN LINK PROTEIN PRECURSOR [H.sapiens]
209	81.82	22	18	X97229	KIR2DL4	killer cell immunoglobulin-like receptor, two domains, long cytoplasmic tail, 4
210	81.82	22	18	D87465	KIAA0275	KIAA0275 gene product

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211	81.82	11	9	S77410	AGTR1	angiotensin receptor 1
212	81.82	11	9	U52112	ARD1	N-acetyltransferase, homolog of <i>S. cerevisiae</i> ARD1
213	81.25	16	13	U18468	PSG4	pregnancy specific beta-glycoprote in 4
214	80	20	16	D29990	SLC7A2	solute carrier family 7 (cationic amino acid transporter, y ⁺ system), member 2
215	80	15	12	M23323	CD3E	CD3E antigen, epsilon polypeptide
216	78.95	19	15	M30607	ZFY	zinc finger protein, Y-linked
217	78.95	19	15	M98833	FLI1	Friend leukemia virus integration 1
218	77.27	22	17	X67292	IGHM	immunoglobulin heavy constant mu
219	76.92	13	10	AA543086		Homo sapiens cDNA: FLJ23270 f1s, clone COL10309, highly similar to HSU33271 Human normal keratinocyte mRNA
220	75	20	15	L13258	SLC34A1	solute carrier family 34 (sodium phosphate), member 1
221	75	12	9	D00174	SERPINF2	serine (or cysteine) proteinase inhibitor, clade F (alpha-2 antiplasmin, pigment epithelium derived factor), member 2
222	73.68	19	14	U05227	SEC4L	GTP-binding protein homologous to <i>Saccharomyces cerevisiae</i> SEC4
223	73.33	15	11	U79240		Human PAS-serine/threonine kinase mRNA, partial cds
224	73.33	15	11	X07994	LCT	lactase
225	72.73	22	16	M80462	CD79A	CD79A antigen (immunoglobulin-associated alpha)
226	71.43	21	15	J04132	CD3Z	CD3Z antigen, zeta polypeptide (TcR/CD3 complex)
227	70.59	17	12	U97145	GFRA2	GNDF family receptor alpha 2
228	69.23	13	9	U05321	SLC16A2	solute carrier family 16 (monocarboxylic acid transporters), member 2 (putative transporter)
229	69.23	13	9	M24405	TCF3	transcription factor 3 (E2A immunoglobulin enhancer binding factors E12/E47)
230	69.23	13	9	U02619	GTF3C1	general transcription factor III C, polypeptide 1 (alpha subunit, 220kDa)
231	68.75	16	11	J04599	BGN	biglycan
232	68.42	19	13	X80818	GRM4	glutamate receptor, metabotropic 4

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233	68.42	19	13	X70991	NAB2	NGFI-A binding protein 2 (ERG1 binding protein 2)
234	68.42	19	13	J00269	KRT6A	keratin 6A
235	68.18	22	15	AA778161	RPL26	ribosomal protein L26
236	66.67	18	12	AI123516		ESTs
237	66.67	15	10	U34623	CD6	CD6 antigen
238	66.67	12	8	AA421322	IGL@	immunoglobulin lambda locus
239	66.67	12	8	M91196	ICSBP1	interferon consensus sequence binding protein 1
240	66.67	12	8	M12807	CD4	CD4 antigen (p55)
241	66.67	12	8	D52745	KIAA0821	lectomedin-2
242	64.29	14	9	S82807	TSHR	thyroid stimulating hormone receptor
243	64.29	14	9	AI027554	DKFZP586J1624	DKFZP586J1624 protein
244	63.64	22	14	U52682	IRF4	interferon regulatory factor 4
245	63.64	22	14	X03066	HLA-DOB	major histocompatibility complex, class II, DO beta
246	63.64	22	14	M15800	MAL	mal, T-cell differentiation protein
247	63.64	22	14	AI366242		ESTs
248	63.64	22	14	X54101	GNLY	granulysin
249	63.64	22	14	T04932		Homo sapiens cDNA: FLJ21545 f s, clone COL06195
250	63.64	22	14	AI248183	PAX5	Paired box gene 5 (B-cell lineage specific activator protein)
251	63.64	11	7	AF064804	SUPT3H	suppressor of Ty (S.cerevisiae) 3 homolog
252	63.64	11	7	D28769	PBX2	pre-B-cell leukemia transcription factor 2
253	63.64	11	7	AA620287		ESTs
254	61.9	21	13	X62071	CDK2	cyclin-dependent kinase 2
255	61.9	21	13	AI271678		ESTs
256	61.9	21	13	X82240	TCL1A	T-cell leukemia/lymphoma 1A
257	61.54	13	8	X78677	KHK	ketoheokinase (fructokinase)
258	61.11	18	11	M75106	CPB2	carboxypeptidase B2 (plasma)
259	59.09	22	13	M74161	INPP5B	inositol polyphosphate-5-phosphatase, 75kD
260	59.09	22	13	AI214175	KIAA0655	huntingtin interacting protein-related
261	58.82	17	10	X15218	SKI	v-ski avian sarcoma viral oncogene homolog
262	57.89	19	11	AA252866	KIP2	DNA-dependent protein kinase catalytic subunit-interacting protein 2
263	57.14	21	12	L19711	DAG1	dystroglycan 1 (dystrophin-associated glycoprotein 1)

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264	55.56	18	10	AA648810	VCP	Valosin-containing protein
265	55	20	11	L13203	FOXI1	forkhead box I1
266	55	20	11	U14534	NR1H2	nuclear receptor subfamily 1, group H, member 2
267	54.55	22	12	M29696	IL7R	interleukin 7 receptor
268	54.55	22	12	M14745	BCL2	B-cell CLL/lymphoma 2
269	54.55	22	12	AI341482	RNB6	RNB6
270	54.55	11	6	U79255	APBA2	amyloid beta (A4) precursor protein-binding, family A, member 2 (X11-like)
271	54.55	11	6	U22526	LSS	lanosterol synthase (2,3-oxidosqualene-lanosterol cyclase)
272	54.55	11	6	S59049	RGS1	regulator of G-protein signalling 1
273	54.55	11	6	M83651	GALGT	UDP-N-acetyl-alpha-D-galactosamine:(N-acetylneuraminyl)-galactosylglucosylceramide N-acetylglucosaminyltransferase (GalNAc-T)
274	53.85	13	7	L02867	HUMPPA	paraneoplastic antigen A
275	53.85	13	7	M35533	LBP	lipopolysaccharide-binding protein
276	53.85	13	7	AI015930	STMN3	Stathmin-like 3
277	53.85	13	7	U11276	KLRB1	killer cell lectin-like receptor subfamily B, member 1
278	53.33	15	8	M89957	CD79B	CD79B antigen (immunoglobulin-associated beta)
279	52.63	19	10	AL009179	H3FK	H3 histone family, member K
280	52.38	21	11	D89618	KPNA3	karyopherin alpha 3 (importin alpha 4)
281	50	22	11	M87790	IGL λ	immunoglobulin lambda locus
282	50	22	11	X72475	IGKC	immunoglobulin kappa constant
283	50	22	11	AF037261	SCAM	vinexin beta (SH3-containing adaptor molecule)
284	50	22	11	M17016	GZMB	granzyme B (granzyme 2, cytotoxic T-lymphocyte-associated serine esterase 1)
285	50	22	11	AA813912	KIAA0130	KIAA0130 gene product
286	50	22	11	AI366182	ESTs	ESTs
287	50	20	10	L31801	SLC16A1	solute carrier family 16 (monocarboxylic acid transporters), member 1
288	50	20	10	D26309	LIMK1	LIM domain kinase 1
289	50	18	9	X68149	BLR1	Burkitt lymphoma receptor 1, GTP-binding protein
290	50	14	7	D86479	AEBP1	AE-binding protein 1

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291	50	14	7	M88338	MSE55	serum constituent protein
292	50	14	7	U73036	IRF7	interferon regulatory factor 7
293	50	14	7	AI336233		ESTs, Weakly similar to carnitine /acylcarnitine translocase [H.sapiens]
294	50	12	6	M19713	TPM1	tropomyosin 1 (alpha)
295	50	12	6	AJ002309	SYNGR3	synaptogyrin 3
296	50	12	6	M73531	RDS	retinal degeneration, slow (retinitis pigmentosa 7)

Accession numbers and gene symbols were retrieved from the Unigene Databases (build#131).

Confirmation by semi-quantitative RT-PCR

To confirm the reliability of the expression differences indicated by microarray analysis, semi-quantitative RT-PCR experiments were performed for the 11 highly up-regulated genes in all of the informative samples (RNASE3, CTSG, MMP9, HP, HPR, H3FJ, HBZ, PLAU, KIAA1254, and two ESTs (Accession No. H23213 and H48537). A 3-μg aliquot of aRNA from each sample was reverse-transcribed for single-stranded cDNAs using random primer (Roche) and Superscript II (Life Technologies, Inc.). Each cDNA mixture was diluted for subsequent PCR amplification with the same primer sets that were prepared for the target DNA- or β-actin-specific reactions. The primer sequences are listed in Table 2. Expression of β-actin served as an internal control. PCR reactions were optimized for the number of cycles to ensure product intensity within the linear phase of amplification. The RT-PCR results were highly concordant to those of the microarray analysis in the great majority of the tested cases (*see* Fig. 1). These data verified the reliability of our strategy to identify commonly up-regulated genes in CML cells.

Table 2 Primer sequences for semi-quantitative RT-PCR experiments

CML Assignment	Accession No.	Symbol	Forward primer	SEQ.ID. NO.	Reverse primer	SEQ.I D.NO.
1	X16545	RNASE3	5'-GTTCCAAAA CTGTTCACTTCC C-3'	No.1	5'-GGTATGGAGA CTGATGAGGACA G-3'	No.2
5	M16117	CTSG	5'-CTTCTGCTGG CCTTTCTCCTA C-3'	No.3	5'-TGTGGACGTT TATTAAGGCTCT G-3'	No.4

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7	J05070	MMP9	5'-GAACCAGCT GTATTTGTTCA AGG-3'	No.5	5'-AAAACAAAGG TGAGAAGAGAG GG-3'	No.6
8	K03431	HPR	5'-TCCTGAATGT GAAGCAGTATG TG-3'	No.7	5'-AGCCTTGCAT TAGTTCTCAGCT A-3'	No.8
15	H23213	EST	5'-GTCCCAAGA TGCATATTTTCC T-3'	No.9	5'-CCGAGCCCAT TAATACTGATAG A-3'	No.10
16	Z98744	H3FJ	5'-ACTTTCTGAC TTAGGCCACAG GT-3'	No.11	5'-ACAGAGTGCT CAGTTCTTCCGT A-3'	No.12
17	M24173	HBZ	5'-TCTCTGACCA AGACTGAGAGG AC-3'	No.13	5'-GAGGATACGA CCGATAGGAACT T-3'	No.14
18	X02419	PLAU	5'-CAGTCACAC CAAGGAAGAG AATG-3'	No.15	5'-CAGTGAGGAT TGGATGAACTAG G-3'	No.16
19	H48537	EST	5'-GTGTGATTAT CAAAAGGGAGT GG-3'	No.17	5'-AATAGTGCCT ATTTAAGGCCG- 3'	No.18
20	AA19144 9	KIAA12 54	5'-TCCTACTTTG GCCAAGTTTGT T-3'	No.19	5'-ACTAAGCTGG TACATGGAATGG A-3'	No.20
57	K01763	HP	5'-AAGGAGATG GAGTGTACACC TTAAA-3'	No.21	5'-TGATTGACTC AGCAATGCAGG- 3'	No.22
	V00478	ACTB	5'-CATCCACGA AACTACCTTCA ACT-3'	No.23	5'-TCTCCTTAGA GAGAAGTGGGG TG-3'	No.24

Industrial Applicability

The gene-expression analysis of CML described herein, obtained through a combination of laser-capture dissection and genome-wide cDNA microarray, has identified specific genes as targets for cancer prevention and therapy. Based on the expression of a subset of these differentially expressed genes, the present invention provides a molecular diagnostic markers for identifying or detecting CML.

The methods described herein are also useful in the identification of additional molecular targets for prevention, diagnosis and treatment of CML. The data reported herein add to a comprehensive understanding of CML, facilitate development of novel diagnostic strategies, and provide clues for identification of molecular targets for therapeutic drugs and preventative agents. Such information contributes to a more profound understanding of tumorigenesis of CML, and

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provide indicators for developing novel strategies for diagnosis, treatment, and ultimately prevention of CML.

All patents, patent applications, and publications cited herein are incorporated by reference in their entirety. Furthermore, while the invention has been described in detail and with reference to specific embodiments thereof, it will be apparent to one skilled in the art that various changes and modifications can be made therein without departing from the spirit and scope of the invention.

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